

09/869206

JORDAN AND HAMBURG LLP
122 East 42nd Street
New York, NY 10168
Tel: (212) 986-2340
Fax: (212) 953-7733

Docket No. **F-7035**Date June 25, 2001

"EXPRESS MAIL" MAILING LABEL NO. EL 666237128 US

DATE OF DEPOSIT June 25, 2001

I hereby certify that this paper or fee is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 CFR 1.10 on the date indicated above and is addressed to the Assistant Commissioner for Patents, Washington, D. C. 20231.

Madeline Gonzalez

(Typed or printed name of person mailing paper or fee)

(Signature of person mailing paper or fee)

THE ASSISTANT COMMISSIONER FOR PATENTS
Washington, D. C. 20231

[] ATTN: BOX PATENT APPLICATION

[X] ATTN: BOX PCT

[X] THIS IS THE NATIONAL STAGE OF PCT/EP99/10334 FILED December 22, 1999

Transmitted herewith for filing is the [X] Utility [] Design patent application of:

Inventor/Application Identifier: **Andrea CZAKI et al.**

For: **AFFINITY SENSOR FOR DETECTING SPECIFIC MOLECULAR BINDING EVENTS AND
USE THEREOF**

Enclosed are:

- [X] 2 sheets of drawings ([X] formal [] informal size A4).
[X] 33 pages of specification, including claims and abstract.
[X] 35 total pages

[X] Combined Declaration/Power of Attorney

[X] Newly executed

[] Copy from prior application

[] Inventors deleted; see attached statement

[] Inventor Information Sheet

[] Incorporation By Reference. The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein

[] Sequence Listing

[] Computer Readable Copy

[] Paper copy

[] The undersigned hereby affirms that the content of the paper and computer readable copies of the Sequence Listing are the same.

[] Cancel in this application original claims _____ of the prior application before calculating the filing fee.

CLAIMS FILED

For	Number Filed	Number Extra	Rate	Basic Fee \$860.00
Total Claims	<u>39</u>	<u>19</u> (over 20) x	\$18.00	<u>342.00</u>
Independent Claims	<u>1</u>	<u>0</u> (over 3) x	\$80.00	
[X] Multiple Dependent Claim			\$270.00	<u>270.00</u>
[X] Reduce by 50% for Small Entity				<u>736.00</u>
[] Foreign Language Filing Fee			\$130.00	
TOTAL FILING FEE				<u>\$736.00</u>
[X] Please charge Deposit Account No. 10-1250 in the amount of A duplicate copy of this sheet is attached.				<u>\$736.00</u>
[X] Please charge to Deposit Account No. 10-1250 any further fees under 37 CFR 1.16; 37 CFR 1.17; 37 CFR 1.492.				

☒ Return Receipt Postcard

☒ Preliminary Amendment

☒ Assignment to (1) INSTITUT FÜR PHYSIKALISCHE HOCHTECHNOLOGIE E.V.

(2) GENETRIX B. V. I. O.

☐ Assignment is of record in prior application Serial No. _____.

☒ Assignment Recordation Form Cover Sheets.

☒ Charge \$80.00 to Deposit Account No. 10-1250 for recording the Assignments.

☒ Information Disclosure Statement and/or Information Disclosure Citation

☐ English translation

☒ Small Entity Status is asserted

☐ Applicant hereby claims the benefit of the filing date of the following provisional application(s) under the provisions of 35 USC 119.

☒ Applicant hereby claims the benefit of the filing date of the following applications under the provisions of 35 USC 119 of which certified copies ☐ will follow ☐ are enclosed ☒ have been filed in the International Bureau ☐ were filed in prior application No. _____.


German Appln. No. 198 60 547.1, filed December 23, 1998

☐ This is a ☐ Continuation ☐ Divisional ☐ Continuation-in-Part of prior application Serial No. _____.

☐ Amend the specification by inserting before the first line the sentence:

--This is a ☐ continuation, ☐ division, ☐ continuation-in-part, of application Serial No. _____, filed _____.

JORDAN AND HAMBURG LLP

By 
C. Bruce Hamburg
Reg. No. 22,389
Attorney for Applicant

F-7035

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant : Andrea CZAKI et al.
Serial No. : Not yet known (U.S. National Stage of
PCT/EP99/10334 filed December 22, 1999)
Filed : Concurrently herewith
For : AFFINITY SENSOR FOR DETECTING SPECIFIC
MOLECULAR BINDING EVENTS AND USE
THEREOF

Assistant Commissioner for Patents
Washington, D.C. 20231

PRELIMINARY AMENDMENT

Sir:

Preliminary to examination, please amend the above-identified patent application as follows:

IN THE TITLE:

Please change the title to read --AFFINITY SENSOR FOR DETECTING
SPECIFIC MOLECULAR BINDING EVENTS AND USE THEREOF.--

IN THE CLAIMS:

Amend claims 1-36 as follows:

1. (Amended) Affinity sensor for detecting specific binding events,
comprising a carrier substrate provided with at least two
electrodes and having a predetermined area, said electrodes

being equidistantly spaced apart from each other and engagingly bordering said area from both sides, at least said area being adapted for receiving immobilized specific binding partners, said specific binding partners being capable of coupling complementarily associated binding partners directly or via further specific binding molecules, said area having a minimum width adapted for capture of at least one complementarily associated being partner provided with one electrically conductive particle within said area in such a way as to allow for formation of a respective tunnel contact junction between the particle and the electrodes.

2. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, wherein said width is under 800 nm.
3. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, wherein the immobilized specific binding partners covers the electrodes with a thickness which permits tunnel effects.
4. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, wherein the electrodes are each two micro-electrodes arranged in a pair, the electrodes being

connected to an amplifier circuit with an associated measuring and evaluating unit so that an electric current flow across the area can be detected when there is a voltage applied across the electrodes.

5. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 4, wherein the electrodes are part of the amplifier circuit and project from out of the latter.
6. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 5, wherein the amplifier circuit is a component of a microchip.
7. (Amend) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, wherein the electrodes are comb-like structures opposingly meshed, whereby there are located affinity areas at least between the respective opposing electrodes.
8. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 7, wherein the comb-like electrodes and the affinity areas are arranged on a common chip surface.

9. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 8, wherein the chip surface is formed by a silicon wafer.
10. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 8, wherein the chip surface is formed by a glass target.
11. (Amended) Affinity sensor for detecting specific molecular binding events as claimed claim 7, wherein the comb-like electrodes are arranged in geometrical symmetry to interdigital structures and a plurality of affinity areas is arranged in a matrix, whereby the electrodes provided outside of the affinity areas are separated from each other at their intersections by an insulating layer arranged between the intersections.
12. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 7, wherein the length of the micro-electrodes is 0.1 mm, the width of the area is 0.1 μm and its effective height is 0.02 μm as well as the affinity areas is at a 1:10 ratio relative to the chip surface.
13. (Amended) Affinity sensor for detecting specific molecular binding events as claim 7, wherein in addition to the affinity areas at

least one reference area is provided which carries inactive binding partner for a reference measurement instead of the specific binding partners.

14. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 7, wherein the occupation density of the specific binding partners on the individual affinity areas is different.
15. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 7, wherein the individual affinity areas carry different specific binding partners.
16. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, 13, 14 or 15, wherein a plurality of reference areas is provided being occupied with different inactive binding partner.

17. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, wherein the specific binding partners enter into coordination compounds.
18. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, wherein the specific binding partners are bioactive or biomimetic molecules.
19. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 17, wherein the specific binding partners are nucleic acids.
20. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 17, wherein the specific binding partners are proteins.
21. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 17, wherein the specific binding partners are saccharides.

22. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, wherein the conductive particles are of sizes in the range of 0.1 μm to 5 μm .
23. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, wherein the conductive particles are of sizes in the nanometer range.
24. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, wherein the conductive particles consist of metal-cluster compounds.
25. (Amended) Method of using the affinity sensor for detecting specific molecular binding events as claimed in claim 1, wherein the affinity sensor is utilized for the detection of complementarily associated binding partners in the form of complex compounds.
26. (Amended) Method according to claim 25, wherein the affinity sensor is utilized for the detection of complementarily associated binding partners in the form of bioactive and biomimetical molecules.

F-7035

27. (Amended) Method according to claim 25, wherein the affinity sensor is utilized for the detection of complementarily associated binding partners in the form of nucleic acids.
28. (Amended) Method according to claim 25, wherein the affinity sensor is utilized for the detection of complementarily associated binding partners in the form of proteins.
29. (Amended) Method according to claim 25, wherein the affinity sensor is utilized for the detection of complementarily associated binding partners in the form of saccharides.
30. (Amended) Method according to claim 24, wherein the affinity sensor is utilized for biomonitoring.
31. (Amended) Method according to claim 30, wherein the affinity sensor is utilized for the detection of cells.

F-7035

32. (Amended) Method according to claim 30, wherein the affinity sensor is utilized for the detection of microorganisms.
33. (Amended) Method according to claim 30, wherein the affinity sensor is utilized for the detection of genetic and microbic diseases.
34. (Amended) Method according to claim 30, wherein the affinity sensor is utilized for the detection of gene expression.
35. (Amended) Method according to claim 32, wherein the affinity sensor is utilized for the detection of microorganisms in ecological populations.
36. (Amended) Method according to claim 30, wherein the affinity sensor is utilized for medical diagnostics.

The changes in the claims are shown by brackets and underscoring in the Appendix hereto.

F-7035

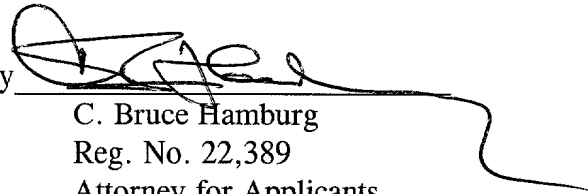
Remarks

This is to place the claims in better condition for examination as to formalities as well as to conform the title to the Combined Declaration/Power of Attorney.

Respectfully submitted,

Jordan and Hamburg LLP

By

A handwritten signature in black ink, appearing to read 'C. Bruce Hamburg', is written over a horizontal line. The signature is stylized with a large initial 'C' and a long, sweeping horizontal stroke extending to the right.

C. Bruce Hamburg
Reg. No. 22,389
Attorney for Applicants

122 East 42nd Street
New York, New York 10168
(212) 986-2340

APPENDIX

AMENDED CLAIMS WITH AMENDMENTS THEREIN INDICATED BY
BRACKETS AND UNDERSCORING

1. (Amended) Affinity sensor for detecting specific binding events, comprising a carrier substrate [(1),] provided with at least two electrodes [(2)] and [a range (4)] having a predetermined area, said electrodes being equidistantly spaced apart from each other and engagingly [border] bordering said [range 4] area from both sides, at least said [range (4)] area being adapted for receiving immobilized specific binding partners [(5)], said specific binding partners [(5)] being capable of coupling complementarily associated binding partners [(6)] directly or via further specific binding molecules [(7)], said [range (4)] area having a minimum width [b being] adapted [to] for capture of at least one complementarily associated being partner [(6), which is] provided with one electrically conductive particle [(62) in] within said [range] area in such a way as to allow for [the] formation of a respective tunnel contact junction between the particle [(62)] and the electrodes [(2)].

F-7035

2. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, [characterized in that the range (4) is given a width b of] wherein said width is under 800 nm.
3. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, [characterized in that] wherein the immobilized specific binding partners [(5) covering also] cover the electrodes [(2)] with a thickness which permits tunnel effects.
4. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, [characterized in that] wherein the electrodes [(2)] are each [designed as] two micro-electrodes [(21) each,] arranged in [pairs] a pair, [which are] the electrodes being connected to an amplifier circuit [(8)] with an associated measuring and evaluating unit [(3)] so that an electric current flow across the [range (4)] area can be detected when there is a voltage applied across the electrodes [(2)].
5. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in [claims 1 and] claim 4, [characterized in that] wherein the electrodes [(2)] are part of the amplifier circuit [(8 projecting] and project from out of the latter.

6. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in [claims 1 and] claim 5, [characterized in that] wherein the amplifier circuit [(8)] is a component of a microchip [(9)].
7. (Amend) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, [characterized in that] wherein the electrodes [(2)] are [designed as] comb-like [meshing] structures oppositely meshed, whereby there are located affinity areas [(41)] at least between the respective opposing [comb-type] electrodes [(22)].
8. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in [claims 1 and] claim 7, [characterized in that] wherein the [comb-type] comb-like electrodes [(22)] and the affinity areas [(41)] are arranged on a common chip surface [(42)].
9. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in [claims 1 and] claim 8, [characterized in that] wherein the chip surface [(42)] is formed by a silicon wafer.
10. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in [claims 1 and] claim 8, [characterized in

that] wherein the chip surface [(42)] is formed by a glass target.

11. (Amended) Affinity sensor for detecting specific molecular binding events as claimed [in claim 1 and] claim 7, [characterized in that] wherein the [comb-type] comb-like electrodes [(22)] are arranged in geometrical symmetry to [the] interdigital structures and [in that] a plurality of affinity areas [(41)] is arranged in a matrix, whereby the electrodes [(2)] provided outside of the affinity areas [(41)] are separated from each other at their intersections [(23)] by an insulating layer [(24)] arranged between the intersections.
12. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in [claims 1 or] claim 7, [characterized in that] wherein the length of the micro-electrodes [(21)] is 0.1 mm, the width [b] of the [range (4)] area is 0.1 μm and its effective height is 0.02 μm as well as the affinity areas [(41)] is at a 1:10 ratio relative to the chip surface [(42)].
13. (Amended) Affinity sensor for detecting specific molecular binding events as [claims 1 or] claim 7, [characterized in that] wherein in addition to the affinity areas [(41)] at least one reference area [(43)] is provided which carries inactive binding

partner [(51)] for a reference measurement instead of the specific binding partners [(5)].

14. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in [claims 1 or] claim 7, [characterized in that] wherein the occupation density of the specific binding partners [(5)] on the individual affinity areas [(41)] is different.
15. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in [claims 1 or] claim 7, [characterized in that] wherein the individual affinity areas [(41)] carry different specific binding partners [(5)].
16. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in [the claims] claim 1, 13, 14 or 15, [characterized in that] wherein a plurality of reference areas [(43)] is provided being occupied with different inactive binding partner [(51)].

17. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, [characterized in that] wherein the specific binding partners [(5)] enter into [co-ordination] coordination compounds.
18. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, [characterized in that] wherein the specific binding partners [(5)] are bioactive or biomimetic molecules.
19. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in [claims 1 and] claim 17, [characterized in that] wherein the specific binding partners [(5)] are nucleic acids.
20. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in [claims 1 and] claim 17, [characterized in that] wherein the specific binding partners [(5)] are proteins.

21. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in [claims 1 and] claim 17, [characterized in that] wherein the specific binding partners [(5)] are saccharides.
22. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, [characterized in that] wherein the conductive particles [(62)] are [defined to a size] of sizes in the range of 0.1 μm to 5 μm .
23. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, [characterized in that] wherein the conductive particles [(62)] are [defined to a size] of sizes in the nanometer range.
24. (Amended) Affinity sensor for detecting specific molecular binding events as claimed in claim 1, [characterized in that] wherein the conductive particles [(62)] consist of metal-cluster compounds.

25. (Amended) [Application] Method of using the affinity sensor for detecting specific molecular binding events as claimed in [one of the preceding claims] claim 1, [characterized in that] wherein the affinity sensor is utilized for the detection of complementarily associated binding partners [(6)] in the form of complex compounds.
26. (Amended) [Application of the affinity sensor for detecting specific molecular binding events as claimed in] Method according to claim 25, [characterized in that] wherein the affinity sensor is utilized for the detection of complementarily associated binding partners [(6)] in the form of bioactive and biomimetical molecules.
27. (Amended) [Application of the affinity sensor for detecting specific molecular binding events as claimed in] Method according to claim 25, [characterized in that] wherein the affinity sensor is utilized for the detection of complementarily associated binding partners [(6)] in the form of nucleic acids.

28. (Amended) [Application of the affinity sensor for detecting specific molecular binding events as claimed in] method according to claim 25, [characterized in that] wherein the affinity sensor is utilized for the detection of complementarily associated binding partners [(6)] in the form of proteins.
29. (Amended) [Application of the affinity sensor for detecting specific molecular binding events as claimed in] Method according to claim 25, [characterized in that] wherein the affinity sensor is utilized for the detection of complementarily associated binding partners [(6)] in the form of saccharides.
30. (Amended) [Application of the affinity sensor for detecting specific molecular binding events as claimed in one of the preceding claims 1 to] Method according to claim 24, [characterized in that] wherein the affinity sensor is utilized for biomonitoring.

31. (Amended) [Application of the affinity sensor for detecting specific molecular binding events as claimed in] Method according to claim 30, [characterized in that] wherein the affinity sensor is utilized for the detection of cells.
32. (Amended) [Application of the affinity sensor for detecting specific molecular binding events as claimed in] Method according to claim 30, [characterized in that] wherein the affinity sensor is utilized for the detection of microorganisms.
33. (Amended) [Application of the affinity sensor for detecting specific molecular binding events as claimed in] Method according to claim 30, [characterized in that] wherein the affinity sensor is utilized for the detection of genetic and microbic diseases.
34. (Amended) [Application of the affinity sensor for detecting specific molecular binding events as claimed in] Method according

to claim 30, [characterized in that] wherein the affinity sensor is utilized for the detection of gene expression.

35. (Amended) [Application of the affinity sensor for detecting specific molecular binding events as claimed in] Method according to claim 32, [characterized in that] wherein the affinity sensor is utilized for the detection of microorganisms in ecological populations.

36. (Amended) [Application of the affinity sensor for detecting specific molecular binding events as claimed in] Method according to claim 30, [characterized in that] wherein the affinity sensor is utilized for medical diagnostics.

AFFINITY SENSOR FOR DETECTING SPECIFIC MOLECULAR BINDING EVENTS AND APPLICATION THEREOF

BACKGROUND OF THE INVENTION

5 The invention relates to an affinity sensor for detecting specific molecular binding events, as is particularly used in the molecular-biological field, for example, in the medical diagnostics, in the biosensor technology or in the DNA-microarray technology, and application of the same.

10

Biosensors are solid phase measuring devices that are comprised of at least one biological receptor, a transducer and a subsequently connected electronic unit.

The receptor utilizes biologically active reagents such as, for example, 15 antibodies for detecting a specific substance such as, for example, antigens. The transduction of detection events into detectable signals is performed by the transducer, for example, by electrochemical, optical, piezoelectric, or calorimetric methods. Thereby, the coupling of the detection events to the transducer can be carried out indirectly or 20 directly. In the first case, the detection events modulate a process which is detected by the transducer. In the second case, the detection events themselves are recorded by the transducer. The transducer is connected to an electronic unit, for example, to a microprocessor followed by modules for signal detection and evaluation.

25 There are numerous application possibilities for such biosensors operating on the basis of molecular detection. These are, among

others, in the field of detection and concentration analysis of biomolecules, the kinetic and equilibrium analysis of biochemical reactions, the control of fermentation processes, the evaluation of receptor-cell-interactions, the clinical analysis, and the cell detection.

- 5 The detection of the presence of bioactive molecules will be performed in the case of nucleic acids, for example, by hybridization with specific and marked nucleic acid probes. The marking of the probes is achieved by enzymatic inclusion of nucleotides that carry radioisotopes such as, for example, tritium, sulphur-35 or phosphorus-
10 32, non-radioactive molecules such as, for example, digoxigenin or biotin and non-radioactive fluorescent molecules, respectively, such as, for example, fluorescein isothiocyanate or 7-amino-4-methylcumarin-3-acetate or metallic particles such as, for example, gold (Nicholl, D. S. T., 1995: Genetische Methoden, Spektrum
15 Akademischer Verlag Heidelberg, p. 24-27).

- In the case of antigens, such as peptides or proteins, the detection of the presence of bioactive molecules is achieved by specific and marked antibodies. The marking of the antibodies is performed by coupling of radioisotopes such as, for example, iodine-125 or tritium,
20 to tyrosine-residuals and histidine-residuals, respectively, by non-radioactive enzymes, for example, alkaline phosphatase or peroxidase, whereby the enzymatic activity is measured, for example, by the conversion of a colorless product into a colored one, by non-radioactive enzymes, for example, haematin which effects the
25 chemiluminescent reaction of hydrogen peroxide and luminol, by non-radioactive enzymes, for example, luciferase which effects bio-

luminescence by means of phosphorized luciferin, or by metallic particles such as, for example, gold (Liddell, E. and Weeks, I., 1996: Antikörpertechniken, Spektrum Akademischer Verlag Heidelberg, p. 87-107).

- 5 The signals from the various marker-molecules used will be evaluated by radio-chemical or electrochemical methods, by optical, piezoelectric, or calorimetric methods for indicating molecular detection events. Thereby, the size of the marker-molecules which emit single signals will lie in the nanometer range.
- 10 The optical and electrochemical methods for representing molecular binding events are the currently most utilized ones.
The problem of the various optical methods is, that the sensitivity and the spatial resolution of the signals emitted by the individual marker-molecules is too low for many applications, that the binding between
15 two links of a specific molecular binding pair cannot be detected, and that the signals are very often superimposed by an unspecific background. These problems of the image generating methods can only be eliminated for a part by an experimental amplification of the signal or by a computer aided statistical image analyzing method.
- 20 The technical limits of the current automation of the image analyzing on the basis of the chip technology lies in the read-out of the various microarray spots. Most of the available technologies are based on the detection of the fluorescence marked binding pairs, which are held in a specific manner to the surface of the chip, whereby the fluorescence
25 detection is performed by an optical read-out of the reactive centers of the microarrays. The application of fluorescent or chemiluminescent

samples is thereby utilized just as in the conventional methods described hereinbefore and is combined with the CCD-imaging (Eggers, M. et al., 1996: Professional Program Proceedings, in Electro '96. IEEE, New York, NY, USA, 364pp.; Heller, M. J., 1996: IEEE-
5 Engineering-in-Medicine-and-Biology-Magazine 15: 100-104), whereby also here the mentioned problems of the conventional image analyzing occur and a binding between two links of a specific molecular binding pair cannot be detected.

The detection of the presence of bioactive molecules can also be
10 obtained by an electrochemical approach by various methods, apart from the commonly used optical methods.

The measurement of redox potential variations in biomolecules is a well-known possibility, which is accompanied by specific binding events, for example, on enzymes. Thereby, the redox potential
15 variations are measured by way of a single electrode, which is provided with biomolecules, and a reference electrode (Heller, A., 1992: Electrical connection of enzyme redox centers to electrodes, J. Phys. Chem. 96: 3579-3587).

The disadvantage of this method lies in the fact that only one single
20 electronic event occurs for one biomolecular binding event, whereby the variation of the redox state, which is effected, lasts only for a short time, so that the detection of each individual binding event had to take place flash-like. This is not possible. The signal obtained is only cumulative so that rare binding events cannot be detected by this
25 technology.

A further possibility for detecting the presence of bioactive molecules in an electrical way is to use biosensors in the form of special measuring electrodes. Such special measuring electrodes generally are comprised of a (strepto)-avidin-coated electrode, whereby the (strepto)-avidin has the property to specifically bind biotin molecules. In this way it is possible to detect peptides, oligonucleotides, oligosaccharides and polysaccharides as well as lipides which are marked with biotin or biotin-derivatives, respectively to couple these as ligands to the (strepto)-avidin-layer. In the latter case, the biotin molecules are the coupling elements. Generally, these biosensors allow to detect antibody/antigen binding pairs, antibody/partial antigen binding pairs, saccharide/lectin binding pairs, protein/nucleic acids binding pairs, and nucleic acids/nucleic acids binding pairs. The detection of the biochemical events occurring at the special measuring electrode takes place in a similar way to that of the before described technology based on redox systems, namely, by measuring the potential variations across a single electrode compared to a reference electrode (Davis, et al., 1995: Elements of biosensor construction; Enzyme Microb. Technol. 17: 130-1035).

A substantial disadvantage of this conventional biosensor technology is the inherent low sensitivity of the measurements attained across the measuring electrodes that cannot be eliminated in that the ligands in an infinitely great density are bound to the measuring electrode, for example, by use of a dextran layer. Due to the additional deposition of, for example, a dextran layer and due to the spatial arrangement of the ligands, the concentration of ligands on the electrodes is indeed raised

up to the sixfold compared to a ligand single layer, but a detection of rare binding events or even of a binding between two elements of a special molecular binding pair is not possible.

Further known possibilities are:

- 5 - the anchoring of specific antibodies on a semiconductor gate of a field-effect transistor, whereby a variation in the charge distribution and, hence, in the circuit of the field-effect transistor is obtained by the selective binding of antigens to the special antibody layer;
- 10 - the immobilizing of special antibodies on the surface of an optical fiber, whereby measurable optical phenomena such as, for example, interfering waves and surface plasmons appear due to the selective binding of antigens to special antibody layers at the site of intersection between the fiber optics and the liquid;
- 15 - as well as the method of surface plasmon resonance, in which, at a definite angle of incidence of the light, the refractive index of a medium is, due to the selective coupling of antigens, measurably varied at a metal-coated glass body which is provided with specific antibodies (Liddell. E. and Weeks, I., 1996: Antikörpertechniken, Spektrum Akademischer Verlag Heidelberg, p. 156-158).
- 20 The disadvantage of these methods is that rare binding events cannot be detected by these technologies.

At present there are only a few methods available which allow a rapid detection of bindings between molecules at low concentrations or even
25 with single molecule pairs (Lemieux, Bertrand et al., "Overview of DNA chip technology." Molecular Breeding 4: 277-289, 1988),

though the biochemical process of the binding pair formation with biosensors, for example, the hybridization of two nucleotide strands or the binding of antibodies to antigens itself runs very quickly, that is, within the range of seconds; biochips can be provided with binding molecules, for example, with specific oligonucleotides (US 5,445,934) or specific proteins (US 5,077,210) so that a chip technology will be possible (Osborne, J. C., 1994: Genosensors. Conference Record of WESCON/94. Idea/Microelectronics. IEEE, New York, N Y, USA: 434pp.; Eggers, M. D. et al., 1993: Genosensors, microfabricated devices for automated DNA sequence analysis. Proc. SPIE-Int. Soc. Opt. Eng. 1998), by which the presence of definite biomolecules can be detected within a few minutes, for example, the presence of genes by use of specific oligonucleotide probes or antigens by use of specific antibodies, and by which great prospects are indicated in the field of biology or medicine, particularly as concerns genetic investigations (Chee, M. et al. 1996: Accessing genetic information with high-density DNA arrays. Science 274: 610-614).

A very promising approach as concerns the detection of binding events between nucleic acid binding-pairs has recently been given by the utilization of the dielectric relaxation frequencies of the DNA to distinguish between hybridized and non-hybridized samples (Beattie et al. 1993. Clin. Chem. 39: 719-722). The detection of the differences in frequencies, however, requires equipment which still is very expensive and which, moreover, still is far from being utilized as a matter of routine.

Furthermore, there is known another way to electronically distinguish hybridized samples from non- hybridized ones, which consists in determining the speed of the electron movements along the DNA strands (US 5,780,234). This determination is based on the fact that the arrangement of the pi-electron orbits in the double-stranded DNA causes the electrons to move faster in double-stranded DNA, that is, in hybridized DNA than in single-stranded DNA (Lipkin et al., 1995: Identifying DNA by the speed of electrons. Science News 147, 117 pp.). To allow for a determination of these electron movements, the target has to be positioned exactly between two molecules. One of these molecules has to be chemically modified in such a way that it acts as an electron donor and the other one such, that it acts as an electron acceptor, so that there is a flow of electrons via electrodes measurable.

This expensive method has the disadvantage that it limits its application to the detection of single-stranded nucleic acids fragments of a defined length and that it is not suited for further biomolecules.

Furthermore, one of the methods for an electrical detection of particles is known from Bezryadin, A., Dekker, C., and Schmid G., 1997: "Electrostatic trapping single conducting nanoparticles between nanoelectrodes." in Applied Physics Letters 71: 1273-1275, in which nanoparticles are captured in a gap formed by electrodes in that a voltage is applied across the electrodes and the capturing of the particles is detected by way of the flow of the current. In contrast to the binding events of biomolecule pairs there is no specific

biochemical binding of the nanoparticles, but the particle is bound to the electrode gap by the electric field.

There is also known from a work by Braun, E., Eichen, Y., Sivan, U., and Ben-Yoseph, G., 1998: "DNA templated assembly and electrode attachment of a conducting silver wire." in Nature 391: 775-778, that DNA molecules can be held between two microstructurized electrodes and these molecules only exhibited an electric conductivity after having been silver coated, whereby this conductivity has nothing to do with specific biochemical binding events of biomolecule pairs.

Alivisatos, A. P., Johnson, K. P., Peng, X., Wilson, T. E., Loweth, C. J., Bruchez Jr. M., P. and Schulz, P., G., 1996: "Organization of nanocrystal molecules using DNA" in Nature 382: 609-611, generated complexes from short single-stringed DNA-molecules and their complementary single-stringed DNA-molecules marked with gold-particles in solution and deposited these on a TEM-grid with a carbon film for a characterization by electron microscope. An electric characterization, however, of the molecule pair binding did not take place.

SUMMARY OF THE INVENTION

It is an object of the present invention to provide an affinity sensor for detecting specific molecular binding events, which in a rapid, sensitive, specific way detects the presence of molecules in routine operation at low expenditures, in particular the presence of bioactive molecules, as well as to provide for special applications of such an affinity sensor.

According to the invention, the object is realized by the features of the claims. More specifically, an affinity sensor consists of a base on which electrodes are disposed in a spaced apart relation capturing a range that is provided with immobilized specified binding partners, which specifically couple complementarily associated binding partners, whereby said binding partners carry electrically conductive particles, so that there can be formed an electrically conductive contact between the electrodes and in this way the variation of the electrical resistance is detectable, when there is a potential applied across the electrodes, as well as the presence of single or a plurality of complementarily associated binding partners, carrying electrically conductive particles.

DETAILED DESCRIPTION OF THE INVENTION

The invention will be explained hereinafter in more detail by virtue of schematical embodiments under reference to the drawings. There is shown in:

Fig. 1 an affinity sensor for detecting specific molecular binding events,

Fig. 2 a schematical representation of the affinity sensor for detecting specific molecular binding events,

Fig. 3 a cross-sectional view of an embodiment of the affinity sensor for detecting specific molecular binding events,

Fig. 4 a plan view of an embodiment of the affinity sensor in the form of an affinity chip, and

Fig. 5 a sectional view along the plane A-A of the affinity chip represented in Fig. 4.

The affinity sensor for detecting specific molecular binding events shown in Fig. 1 and 2, is comprised of a carrier substrate 1 which is provided with electrodes 2 enclosing a range 4 that is provided with immobilized specific binding partners 5. Thereby the range 4 represents a discontinuity in an electric circuit that includes an amplifier circuit 8, which can be part of a microchip 9, as well as a measuring and evaluating unit 3, whereby in the present example the electrodes 2, which limit the range 4, are associated to the electric circuit and define a minimum width b of the range 4. The specific binding partners 5 are capable of coupling complementarily associated binding partners 6 specifically and directly or via further specific binding molecules 7, whereby the complementarily associated binding partners 6 including electrically conductive particles 62 are directly coupled or via binding molecules. The range 4 is, by the arrangement of the electrodes 2, so dimensioned in its width and effective height that the coupling of the immobilized specific binding partners 5 to the complementarily associated binding partner 6 which carry the electrically conductive particles 62 or, via further specific binding molecules 7, with the complementarily associated binding partners 6 which carry the electrically conductive particles 62. Provided that the specific binding partners 5 are realized by the molecules of a nucleic acid probe species, the complementarily associated binding partners 6, which carry the electrically conductive particles 62, by nucleic acids

and the electrically conductive particles 62 by nanoparticles of a size of 20 nm, then the minimum width b of the range 4 is 25 nm and its effective height 20 nm.

The coupling of the specific binding partners 5 in the range 4 to the complementarily associated binding partners 6 carrying the electrically conductive particles 62 effects, when there is applied a voltage across the electrodes 2 (refer to Fig. 1), the motion of the electrons via the electron transport barrier in such a way that the electrically conductive particles 62 bridge the range 4 so that the electrons tunnel from particle 62 to particle 62 and to the electrodes 2, as a result thereof a permanent variation of the electric resistance across the range 4 between the electrodes 2 can be measured by aid of the post-connected amplifier circuit 8 in combination with the measuring and evaluating unit 3.

The measurements can also be performed in a humid environment, in particular by aid of a gel layer, instead of measuring in a dry state.

In order to enhance the electric conductivity of the range 4 between the electrodes 2, which is achieved by way of the complementarily associated binding partners 6 in cooperation with the electrically conductive particles 62, already known electron-transfer-mediators or effective diffusing electron donors and electron acceptors can be used, such as water soluble ferrocene/ferricinium, hydroquinone/quinone, reducible and oxidizable components from organic salts, cobaltocenes, hexacyanides and octacyanides of molybdenum, tungsten, and iron, respectively, macrocycles and chelating ligands from the transition metals such as cobalt, ruthenium, and nickel, including

Co(ethylenediamine)₃- and Ru(ethylenediamine)₃- and trisbipyridyl- and hexamine-complexes from transition metals such as Co, Ru, Fe, and/respectively, organic molecules such as 4-4'-bipyridines and 4-mercaptopyridines, which are free in solution or present in a gel deposited on the carrier substrate 1 or in a polymer deposited on the carrier substrate 1. When a known gel-based matrix immobilization utilizes nucleic acids as specific binding partners 5 then, due to the three-dimensional structure of the polymer, it exhibits the advantage that a greater number of capturing ligands is immobilized on the small surface section of the range 4. By using a highly porous hydro-gel, the hybridization rate, for example, of the nucleic acids which are the specific binding partners 5 and the complementarily associated binding partners 6, which carry the electrically conductive particles 62, is increased and lies within ranges as they are known for nucleic acids in solution.

The affinity sensor shown in Fig. 3 and 4, which is in the form of a affinity chip, is characterized in that the electrodes 2 are designed as micro-electrodes 21, which are arranged in two pairs each, capturing a respective affinity area 41. Thus, a matrix of affinity areas 41 results, which is adapted to simultaneously and electrically detect in the different interspaces 4 a plurality of various couplings.

Thereby, the individual affinity areas 41 are designed in an inter-digital electrode structure arranged upon a chip surface 42. The chip surface 42 consists of silicon or glass upon which, for example, a dielectric oxide layer is provided. Due to the digitally branched micro-electrodes 21, which, for example, can be manufactured to yield the

shape of comb-like electrodes 22, the ranges 4 on the affinity area 41 can be defined to have a length within a range of 20 μm . The micro-electrodes 21 are spaced apart and electrically separated from each other by an interposed insulating layer 24, as shown in Fig. 5, which is provided at the intersections 23 of the micro-electrodes 21. Thereby and provided that the specific binding partners 5 are realized by the molecules of a nucleic acid probe species, the complementarily associated binding partners 6, which carry the electrically conductive particles 62, are nucleic acids and the electrically conductive particles 62 are nanoparticles of a size of 20 nm, then the ranges 4 have an effective height of 100 nm and a width of 200 nm. Consequently, at least one coupling, which establishes a contact between the micro-electrodes 21, is achieved between the immobilized specific binding partners 5 and the complementarily associated binding partners 6 which carry the electrically conductive particles 62. In this example, the immobilized specific binding partners 5 are capturing ligands in the form of nucleic acid probes and the complementarily associated binding partners 6, which carry the electrically conductive particles 62, are target molecules in the form of nucleic acids. The oligonucleotide probes immobilized as specific binding partners 5 are bound to the silanized carrier substrate 1 via an amino group, whereby a probe density in an order of size of 10,000 molecules per μm^2 is attained in this example. The complementarily associated binding partners 6 are oligonucleotides in this example, which are marked with gold particles, the hybridization conditions depending on the respectively used probes.

Alternatively, the affinity areas 41 can be provided with various immobilized specific binding partners 5 in sectors, which are respectively separated from each other.

Affinity areas 41 with immobilized specific binding partners 5 and
5 reference areas 43 with immobilized inactive binding partners 51 are provided on affinity chips, represented in Fig. 3 and 4, so that the measurement of the electric resistance between the micro-electrodes 21 is carried out as a reference measurement of the electric resistance between an affinity area 41 and a reference area 43, whereby the
10 micro-electrodes 21 can be designed as comb-type electrodes 22. Thereby the immobilized specific binding partners 5 and the immobilized inactive binding partners 51 can be of a thickness which, when covering the electrodes 21, permits the tunnel effect, rendering the manufacture of the chips technologically more easily.

15 Since the reference area 43 is free from immobilized specific binding partners 5, due to the occupation by inactive binding partners 51, this space between the two micro-electrodes 21, insulated from each other, represents an electrical barrier so that there does not take place a measurable electron transfer between them.

20 The affinity area 41, which in contrast thereto carries immobilized specific binding partners 5, binds via the latter and through the coupling event the complementarily associated electrical binding partners 6, which carry the electrically conductive particles 62, so that as a result thereof, by the conductive particles 62, the space of the
25 affinity areas 41 between the micro-electrodes 21, which are designed as comb-type electrodes 22, is divided into a plurality of gaps of

nanometer width. The nano-gaps formed by the electrically conductive particles 62 result in that an electron transfer is possible between the two contact faces of the micro-electrodes 21 by virtue of the tunnel effect, so that the variation of the resistance can be detected via the amplifier circuit 8 by means of a measuring and evaluating unit 3,
5 when there is a voltage applied across the micro-electrodes 21. In the present example, the voltage applied lies in an order of size of less than one volt.

Alternatively to the measurement of the potential applied across the affinity 41 by an electrode system comprised of reference electrode,
10 sample electrode and counter electrode, it is also possible to employ other methods of an electrical detection such as, for example, potentiometric and voltametric measurements.

Standard chemical linkers such as, for example, amino-modified ligands, are used to immobilize the specific binding partners 5 and the inactive binding partners 51, respectively, such as, for example,
15 antibodies or nucleotide probes, so that the chemical linkers are bound to the silanized chip surface 42 and constitute the affinity areas 41 and the reference areas 43, respectively.

The marking of the complementarily associated binding partners 6
20 such as, for example, protein targets or the target nucleic acid, by means of electrically conductive particles 62 is performed according to the known methods such as, for example, the final marking with marked oligonucleotides, by utilizing ligases.

25

In the following, the manufacturing of affinity sensors according to the present will be described in more detail. In a preferred embodiment the affinity sensor is comprised of a plurality of ranges 4 (also referred to as detection ranges), whereby each of which is captured by at least
5 two electrodes 2. These detection ranges are provided with specific binding partners (capture molecules) 5 such as antibodies, fragments of antibodies or DNA-, RNA- or PNA-oligonucleotides, to which definite associated binding partners (target molecules) 6 bind in a specific manner. The specific binding partners 5 are defined as marked
10 or non-marked molecules, which can be selected for being bound to the desired target molecule in the ranges 4 of the affinity sensor. To this end, not only conventional (bio)molecular binding pairs can be utilized as capturing molecules and as target molecules, but also specific chemical binding pairs as known from the combinatorial
15 chemistry, which can also be utilized as binding pairs within the frame of the invention. The formation of this described specific binding can be understood as a primary binding event. It is possible to carry out the detection of this primary binding in a one-step procedure or in a multi-step procedure, whereby the specific co-immobilization of the
20 material, which transfers the electrons, for example, the gold particles 62, is carried out in the last step each. This co-immobilization can be performed by specific kinds or unspecific kinds of molecular interaction, such as a hybridization of probes marked with gold onto the desired target molecule or by a direct marking of the target
25 molecule with the properties of an electron transfer in such a way that this marking can be electronically detected. The mentioned co-

immobilization is, in principle, separated from the primary binding event, in dependence, however, therefrom and can be performed simultaneously. Thus, the co-immobilization or attachment of material, which transfers electrons, to the designated surface of the affinity sensors can be taken as an indirect result of the primary binding. The detection of this co-immobilization is obtained by an electronic measurement of the variation of the electric conductivity across the measuring range, this variation of the electric conductivity being an indication of the presence of target molecules. The primary binding of electron-transferring material can be exploited to induce secondary depositions which are adapted to transport electrons. It lies within the scope of the present invention that the specific binding of target molecules can be detected by way of a multi-step process, which comprises at least one step by way of which electron-transferring material is deposited, this material effecting a reduction of the electric resistance across the measuring range. It is possible to use organic or inorganic substances or compounds for the electron conductive particles 62. This conductivity is used for detecting and marking of the desired target molecule, that is, for detecting the presence thereof.

In the following and without limiting the present invention thereto there will be described several possibilities of preparation steps for manufacturing an affinity sensor according to the present invention.

A. To prepare the required electrodes, a silicon wafer having on one side an oxide layer of about 1 μm thickness is coated by sputtering with a bonding layer, for example, of 3 nm Ti, to said oxide layer and

a gold layer of a thickness of 50-100 nm. To be able to provide for the electron gap width in the lower nanometer range, a multi-layer masking is utilized for the micro-structuring. To this end, a coating with a carbon (30 nm) is performed, followed by a coating with a metal combination (Ti and NiCr, respectively, of a thickness of 10
5 nm). Subsequently, an electron beam resist (150 nm) is deposited by spinning-on. The exposure is realized by a mix-match-technology, in the course of which the large-area electrodes 2 are generated by means of a shaped-electron-beam exposure device and the minute gaps
10 between the electrodes 2 by means of a point-beam electron-beam exposure device. The structure is transferred to the metal layer by ion-beam etching (IBE) and to the carbon layer by a reactive ion-etching (RIE). The transfer of the structure to the gold-layer and the bonding layer is carried out by way of an IBE-process. Finally, the masking
15 layer is removed in an O₂-RIE-process at a simultaneous surface activation.

In the following, techniques will be described which are based on a silanization of the surface of the chips. Due to this silanization, the
20 surfaces are activated for binding amino-modified oligonucleotides. Two different methods for the silanization and subsequent immobilization will be explained here. Of course, there are also other possibilities for surface activation and immobilization, apart from the silanization.

25

B.1. Silanization by application of 3-aminopropyltrimethoxysilane APTES:

The pre-structured chips with gold electrodes, as described by example under A., are purged in an ultrasonic bath and, in sequence in
5 concentrated nitric acid, in hydrogen peroxide solution (30%) and water, and subsequently dried for 5 minutes at 80°C. Then the chips will be incubated for 2 min. in a 1% silane solution in 95% acetone/water. After having been washed for ten times in acetone for 5 minutes each, the chips will be dried at 110°C. Then they will be
10 incubated for 2 h in a 0.2%-phenylenediisothiocyanate solution in 10% pyridine/dimethylformamide and washed with methanol and acetone. Chips activated in this manner can be stored in a desiccator at 4°C for a longer time.

Subsequently, the linkage of the amino-modified oligonucleotides is
15 performed, to this purpose a drop of the oligonucleotide solution (2mM in 100mM sodium carbonate/sodium bicarbonate buffer) is deposited upon the chip. The parallel application of small drops of different oligonucleotides allows a parallelization, for example, by use of an embodiment of the affinity sensor according to Fig. 4. The
20 deposition of the mentioned drops can be performed by means of micro-pipettes, spotters or other available techniques suited for the application of small amounts of samples. Then, the chips are incubated in a moisture chamber at 37°C for about 1-2 h. After removal of the drops the chips will be washed with 1%-ammonia solution for one-
25 time, and three-times with water. Then drying is carried out at ambient temperature.

B.2. A second possibility of silanization is carried out by application of 3- glycidoxypropyltrimethoxysilane (GOPS), to this end, as described under B1., the chips are purged and subsequently are treated in an ultrasonic bath, each for 12 min. with hexane, acetone and
5 ethanol. Then the chips are dried for 5 minutes at 80°C. The silanization is carried out with 1 mM GOPS in dry toluol at 80°C for 6-8 h. The chips are thoroughly washed with ethyl acetate and are ready for immediate use.

Subsequently, the linkage of the amino-modified oligonucleotides is
10 performed. To this purpose a drop of the oligonucleotide solution (5-50 μ M in 0.1M KOH) is deposited upon the chip and the chip is incubated in the moisture chamber at 37°C for 6 h.. Again a parallelization, as referred to under B.1. can be obtained due to the deposition of a plurality of drops with different oligonucleotides. Then
15 the drops are allowed to dry, and then washing is carried out with water at 50°C under continuous shaking, followed by drying at ambient temperature.

C. In this part of the specification there will be described the
20 possibility of marking oligonucleotide probes with colloidal gold. To start with, there is required a preparation of the thiolated oligonucleotide, which is carried out as follows: the 3'-alkylthiol modified oligonucleotides are solid-phase bound to a dithiol-compound by the manufacturer to protect its functional group. By
25 separation from the carrier material the functional group will be released and is then in the active state. The separation takes place in 50

mM DTT (dithiothreitol) in concentrated ammonium hydroxide at 55°C for 16 h (original solution: 4-8 mg solid-phase bound oligonucleotide, 450 µl water, 50 µl 1M DTT, 50 µl cc ammonium hydroxide). After incubation the liquid phase is separated from the solid phase (Controlled Pored Glass, CPG) and desalinated by way of column chromatography. The oligonucleotides are then washed out in reaction buffers. The concentration of the single chromatography fractions is then detected by a spectrophotometer.

The reaction solution will be incubated at 55°C for 16 h at 600 revolutions per minute in a thermomixer, and then centrifugated for 2-3 min. at an acceleration of about 16,000 m/s². Fractions that are prepared in this manner can be stored for more than 4 weeks at -20°C. The binding of the thiolated oligonucleotides to colloidal gold will be described by example in the following:

There are added to 5 ml gold solution (about 17 nM) 2.5 OD (260 nm) alkylthiololigonucleotides [OD (260 nm) = optical density at 260 nm], (final concentration 3.6 nM). Subsequently to a pre-incubation for 16 h at ambient temperature, incubation is carried out after a setting to 0.1 M NaCl/10mM sodium phosphate buffer (pH 7.0) for 40 h at ambient temperature. Thereafter, again a centrifugation takes place for 25 min. at an acceleration of about 16,000 m/s². The resulting pellet is washed with 5 ml 0.1 M NaCl/10 mM sodium phosphate buffer (pH 7.0), followed by a further centrifugation for 25 min. at an acceleration of 16,000 m/s². The re-dispersion is carried out in 5 ml 0.3 M NaCl/10 mM sodium phosphate buffer (pH 7.0).

40 μ l of the aqueous solution with colloidal gold particles (diameter of 30 nm in the example) obtained in the above described manner are placed in the range 4 between the electrodes 2. After drying, electric measurements, which have been described herein further up, show a linear current-voltage characteristic which is indicative of an ohmic behaviour of the aggregated gold colloids in the range under consideration. A current of 0.3 μ A was measured at a voltage of about 0.3 volt applied across the electrodes 2.

- 10 The affinity sensor as, for example, disclosed in connection with Fig. 3 and 4 and in form of the affinity chips, can find a variety of applications as, for example, in the molecular biology and in the medical diagnostics where specific bindings of bioactive molecules to their corresponding binding partners, for example, DNA, proteins, 15 saccharides are to be determined.

Based on the electrical detection of specific molecular binding events, the affinity sensor allows to perform a bio-monitoring of, for example, molecules, viruses, bacteria, and cells in the most diverse samples, for 20 example, in clinical samples, in samples of food and from the environment such as, for example, from clarification plants, whereby such monitoring is performed in a quick, sensitive and specific way.

LIST OF REFERENCE NUMERALS

	1	carrier substrate
	2	electrode
	21	micro-electrodes
	22	comb-type electrodes
5	23	intersection
	24	insulating layer
	3	measuring and evaluating unit
	4	range
	41	affinity area(s)
10	42	chip surface
	43	reference area(s)
	5	specific binding partners
	51	inactive binding partner(s)
	6	complementarily associated binding
15		partners
	62	electrically conductive particles
	7	specific binding molecules
	8	amplifier circuit
	9	microchip
20	A	plane of section
	b	width of range 4

CLAIMS

1. Affinity sensor for detecting specific binding events comprising a carrier substrate (1), provided with at least two electrodes (2) and a
5 range (4), said electrodes being equidistantly spaced apart from each other and engagingly border said range (4) from both sides, at least said range (4) being adapted for receiving immobilized specific binding partners (5), said specific binding partners (5) being capable of coupling complementarily associated binding
10 partners (6) directly or via further specific binding molecules (7), said range (4) having a minimum width b being adapted to capture at least one complementarily associated binding partner (6), which is provided with one electrically conductive particle (62) in said range in such a way to allow for the formation of a respective
15 tunnel contact junction between the particle (62) and the electrodes (2).
2. Affinity sensor for detecting specific molecular binding events as claimed in claim 1, characterized in that the range (4) is given a
20 width b of under 800 nm.
3. Affinity sensor for detecting specific molecular binding events as claimed in claim 1, characterized in that the immobilized specific binding partners (5) covering also the electrodes (2) with a
25 thickness which permits tunnel effects.

4. Affinity sensor for detecting specific molecular binding events as claimed in claim 1, characterized in that the electrodes (2) are designed as two micro-electrodes (21) each, arranged in pairs, which are connected to an amplifier circuit (8) with an associated measuring and evaluating unit (3) so that an electric current flow across the range (4) can be detected when there is a voltage applied across the electrodes (2).
5. Affinity sensor for detecting specific molecular binding events as claimed in claims 1 and 4, characterized in that the electrodes (2) are part of the amplifier circuit (8) projecting from out of the latter.
6. Affinity sensor for detecting specific molecular binding events as claimed in claims 1 and 5, characterized in that the amplifier circuit (8) is a component of a microchip (9).
7. Affinity sensor for detecting specific molecular binding events as claimed in claim 1, characterized in that the electrodes (2) are designed as comb-like meshing structures, whereby there are located affinity areas (41) at least between the respective opposing comb-type electrodes (22).
8. Affinity sensor for detecting specific molecular binding events as claimed in claims 1 and 7, characterized in that the comb-type electrodes (22) and the affinity areas (41) are arranged on a common chip surface (42).

9. Affinity sensor for detecting specific molecular binding events as claimed in claims 1 and 8, characterized in that the chip surface (42) is formed by a silicon wafer.

5

10. Affinity sensor for detecting specific molecular binding events as claimed in claims 1 and 8, characterized in that chip surface (42) is formed by a glass target.

10 11. Affinity sensor for detecting specific molecular binding events as claimed in claims 1 and 7, characterized in that the comb-type electrodes (22) are arranged in geometrical symmetry to the interdigital structures and in that a plurality of affinity areas (41) is arranged in a matrix, whereby the electrodes (2) provided outside
15 of the affinity areas (41) are separated from each other at their intersections (23) by an insulating layer (24) arranged between the intersections.

12. Affinity sensor for detecting specific molecular binding events as
20 claimed in claims 1 or 7, characterized in that the length of the micro-electrodes (21) is 0.1 mm, the width b of the range (4) is 0.1 μm and its effective height is 0.02 μm as well as the affinity areas (41) is at a 1:10 ratio relative to the chip surface (42).

25 13. Affinity sensor for detecting specific molecular binding events as claimed in claims 1 or 7, characterized in that in addition to the

affinity areas (41) at least one reference area (43) is provided which carries inactive binding partner (51) for a reference measurement instead of the specific binding partners (5).

5 14. Affinity sensor for detecting specific molecular binding events as claimed in claims 1 or 7, characterized in that the occupation density of the specific binding partners (5) on the individual affinity areas (41) is different.

10 15. Affinity sensor for detecting specific molecular binding events as claimed in claims 1 or 7, characterized in that the individual affinity areas (41) carry different specific binding partners (5).

15 16. Affinity sensor for detecting specific molecular binding events as claimed in the claims 1, 13, 14 or 15, characterized in that a plurality of reference areas (43) is provided being occupied with different inactive binding partner (51).

20 17. Affinity sensor for detecting specific molecular binding events as claimed in claim 1, characterized in that the specific binding partners (5) enter into co-ordination compounds.

25 18. Affinity sensor for detecting specific molecular binding events as claimed in claim 1, characterized in that the specific binding partners (5) are bioactive or biomimetic molecules.

19. Affinity sensor for detecting specific molecular binding events as claimed in claims 1 and 17, characterized in that the specific binding partners (5) are nucleic acids.
- 5 20. Affinity sensor for detecting specific molecular binding events as claimed in claims 1 and 17, characterized in that the specific binding partners (5) are proteins.
- 10 21. Affinity sensor for detecting specific molecular binding events as claimed in claims 1 and 17, characterized in that the specific binding partners (5) are saccharides.
- 15 22. Affinity sensor for detecting specific molecular binding events as claimed in claim 1, characterized in that the conductive particles (62) are defined to a size of 0.1 μm to 5 μm .
- 20 23. Affinity sensor for detecting specific molecular binding events as claimed in claim 1, characterized in that the conductive particles (62) are defined to a size in the nanometer range.
24. Affinity sensor for detecting specific molecular binding events as claimed in claim 1, characterized in that the conductive particles (62) consist of metal-cluster compounds.
- 25 25. Application of the affinity sensor for detecting specific molecular binding events as claimed in one of the preceding claims,

characterized in that the affinity sensor is utilized for the detection of complementarily associated binding partners (6) in the form of complex compounds.

- 5 26. Application of the affinity sensor for detecting specific molecular binding events as claimed in claim 25, characterized in that the affinity sensor is utilized for the detection of complementarily associated binding partners (6) in the form of bioactive and biomimetical molecules.

10

27. Application of the affinity sensor for detecting specific molecular binding events as claimed in claim 25, characterized in that the affinity sensor is utilized for the detection of complementarily associated binding partners (6) in the form of nucleic acids.

15

28. Application of the affinity sensor for detecting specific molecular binding events as claimed in claim 25, characterized in that the affinity sensor is utilized for the detection of complementarily associated binding partners (6) in the form of proteins.

20

29. Application of the affinity sensor for detecting specific molecular binding events as claimed in claim 25, characterized in that the affinity sensor is utilized for the detection of complementarily associated binding partners (6) in the form of saccharides.

25

30. Application of the affinity sensor for detecting specific molecular binding events as claimed in one of the preceding claims 1 to 24, characterized in that the affinity sensor is utilized for biomonitoring.

5

31. Application of the affinity sensor for detecting specific molecular binding events as claimed in claim 30, characterized in that the affinity sensor is utilized for the detection of cells.

10 32. Application of the affinity sensor for detecting specific molecular binding events as claimed in claim 30, characterized in that the affinity sensor is utilized for the detection of microorganisms.

15 33. Application of the affinity sensor for detecting specific molecular binding events as claimed in claim 30, characterized in that the affinity sensor is utilized for the detection of genetic and microbic diseases.

20 34. Application of the affinity sensor for detecting specific molecular binding events as claimed in claim 30, characterized in that the affinity sensor is utilized for the detection of gene expression.

25 35. Application of the affinity sensor for detecting specific molecular binding events as claimed in claim 32, characterized in that the affinity sensor is utilized for the detection of microorganisms in ecological populations.

36. Application of the affinity sensor for detecting specific molecular binding events as claimed in claim 30, characterized in that the affinity sensor is utilized for medical diagnostics.

ABSTRACT

The invention relates to an affinity sensor for detecting specific molecular binding events, for use in the field of molecular biology, e.g., in medical diagnostics, especially in biosensor technology or in DNA microarray tests. The aim of the invention is to provide an affinity sensor of this type for rapidly, sensitively, specifically, economically and routinely detecting the presence of molecules, especially bioactive molecules, and to provide special applications for an affinity sensor of this type. To this end, the inventive affinity sensor consists of a support substrate (1) which is provided with at least two electrodes (2). Said electrodes are situated equidistantly from each other and cover an area (4) on both sides, at least this area (4) being provided for receiving immobilised specific binding partners (5) which are capable of coupling complementary corresponding binding partners (6) directly or with other specific binding molecules (7). The area (4) is established with a minimum width b , in such a way that at least one complementary corresponding binding partner (6) which is provided with an electroconductive particle (62) can be received in said area in such a way as to guarantee the possibility of a tunnel-type contact junction forming between the particle (62) and the electrodes (2) in each case. The affinity sensor is used for biomonitoring.

1/2

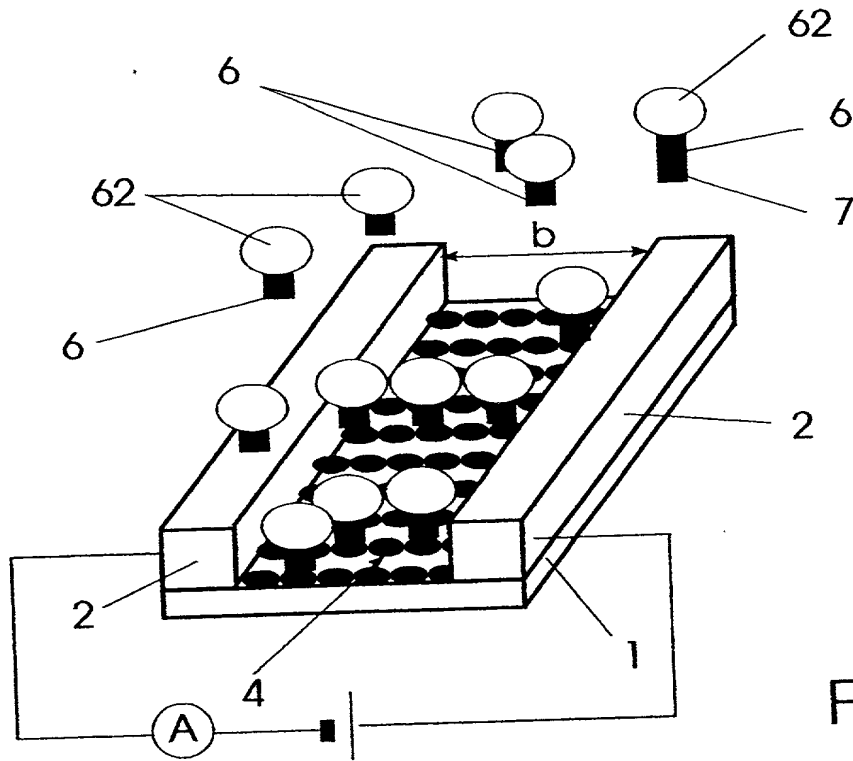


Fig. 1

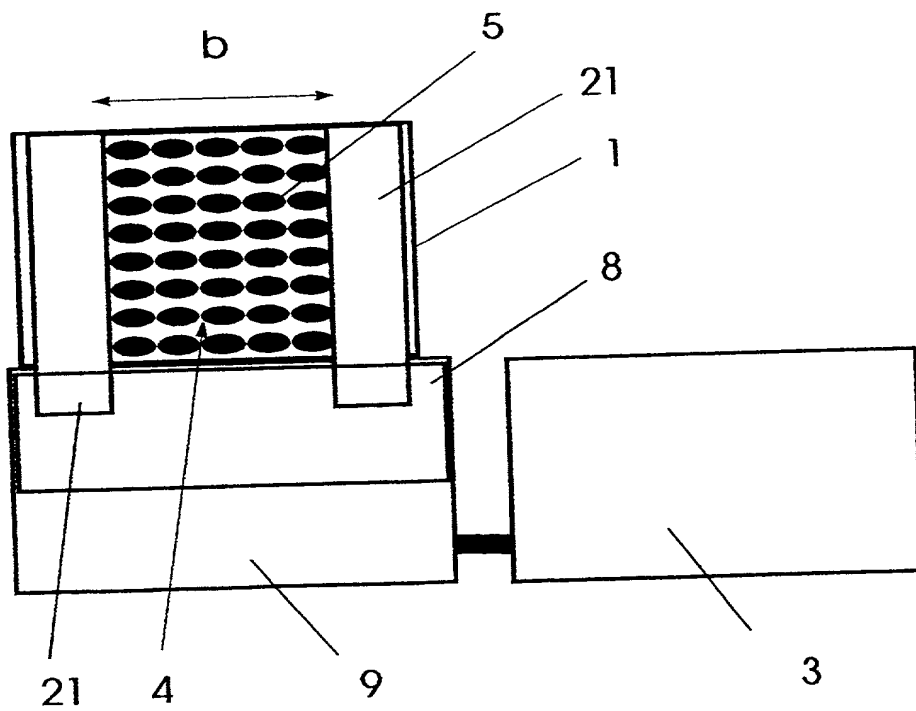


Fig. 2

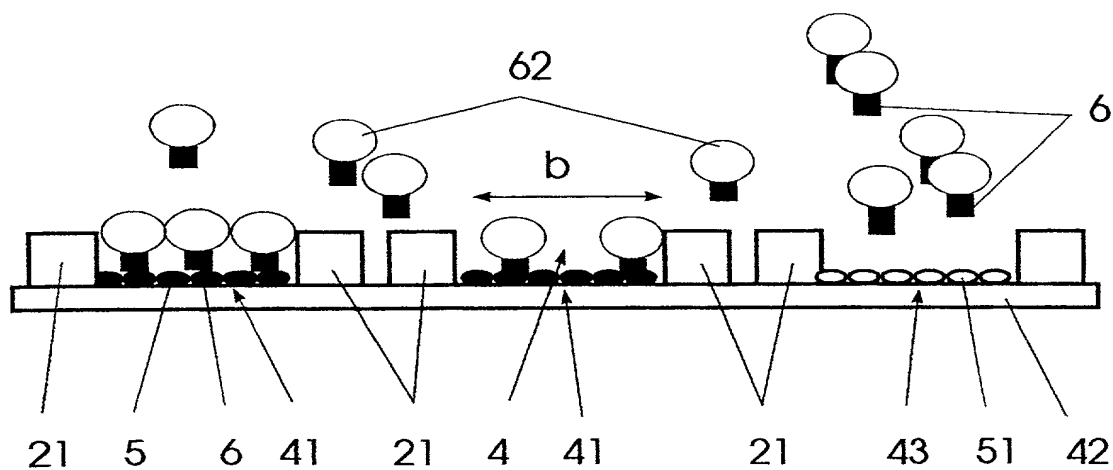


Fig. 3

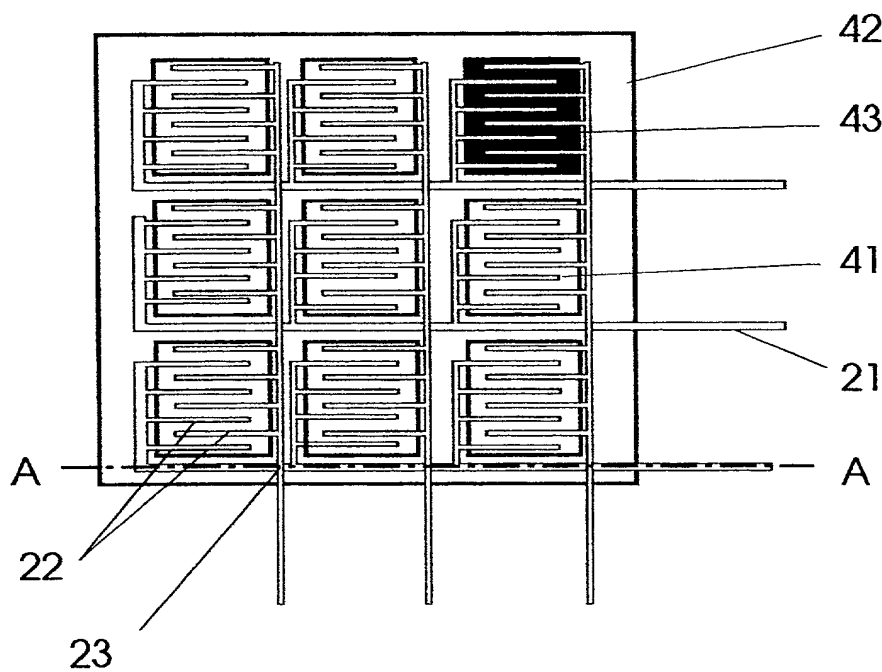


Fig. 4

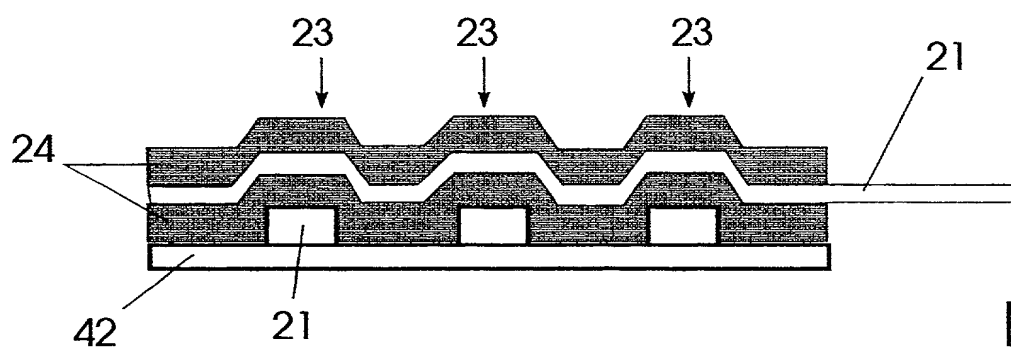


Fig. 5

**COMBINED DECLARATION FOR PATENT APPLICATION AND
POWER OF ATTORNEY**

(Includes Reference to PCT International Applications)

Agency's Docket Number

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

**AFFINITY SENSOR FOR DETECTING SPECIFIC MOLECULAR BINDING
EVENTS AND USE THEREOF**

the specification of which (check only one item below):

☐ is attached hereto.

☐ was filed as United States application

Serial No. _____

on _____

and was amended

on _____

(if applicable).

☒ was filed as PCT international application

Number _____

PCT/EP99/10334

on _____

December 22, 1999

and was amended under PCT Article 19

on _____

(if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, §1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application(s) for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed:

PRIOR FOREIGN/PCT APPLICATION(S) AND ANY PRIORITY CLAIMS UNDER 35 U.S.C. 119:			
Country (if PCT indicate "PCT")	Application Number	Date of Filing (day, month, year)	Priority Claimed Under 35 USC 119
DE	198 60 547.1	23.12.1998	<input checked="" type="checkbox"/> Yes <input type="checkbox"/> No
			<input type="checkbox"/> Yes <input type="checkbox"/> No
			<input type="checkbox"/> Yes <input type="checkbox"/> No
			<input type="checkbox"/> Yes <input type="checkbox"/> No
			<input type="checkbox"/> Yes <input type="checkbox"/> No

COMBINED DECLARATION FOR PATENT APPLICATION AND
POWER OF ATTORNEY (Continued)
(Includes Reference to PCT International Applications)

Attorney's Docket Number

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) or PCT international application(s) designating the United States of America that is/are listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in that/those prior application(s) in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, §1.56(a) which occurred between the filing date of the prior application(s) and the national or PCT international filing date of this application:

PRIOR U.S. APPLICATIONS OR PCT INTERNATIONAL APPLICATIONS DESIGNATING THE U.S. FOR BENEFIT UNDER 35 U.S.C. 120:					
U.S. APPLICATIONS			STATUS (Check One)		
U.S. Application Number	U. S. Filing Date		Patented	Pending	Abandoned
PCT APPLICATIONS DESIGNATING THE U.S.					
PCT Application No	PCT Filing Date	U.S. Serial Numbers Assigned (if any)			

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith.

Frank J. Jordan Reg. No. 20,456
C. Bruce Hamburg Reg. No. 22,389
Lainie E. Dolinger Reg. No. 36,123

Herbert F. Ruschmann Reg. No. 35,341
Marvin Turken Reg. No. 18,330
Alfred D'Andrea Reg. No. 27,752

Send Correspondence To: Jordan and Hamburg
122 East 42nd Street
New York, New York 10168

Direct Telephone Calls to:
C. Bruce Hamburg
(212) 986-2340

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Full Name of Sole or First Inventor	Inventor's Signature	Date
<u>Wolfgang FRITZSCHE</u>	<u>W. Fritz</u>	<u>23.5.01</u>
Residence	Citizenship	
<u>Jena</u>	<u>German</u>	
Post Office Address		
<u>Hufelandweg 16 D-07743 Jena</u> <u>Germany</u>		

Full Name of Second Joint Inventor, if any	Inventor's Signature	Date
<u>Andrea CZAKI</u>	<u>Andrea Czak</u>	<u>23.5.01</u>
Residence	Citizenship	
<u>Camburg</u>	<u>German</u>	
Post Office Address		
<u>Georgstr. 9 D-07774 Camburg</u> <u>Germany</u>		

3-80

Full Name of Sole or First Inventor Johann Michael KÖHLER	Inventor's Signature <i>Johann Köhler</i>	Date 23.5.01
Residence Golmsdorf		Citizenship German
Post Office Address Untergasse 8, D-07751 Golmsdorf Germany		

DEX

4-80

Full Name of Second Joint Inventor, if any Antje WIEGAND	Inventor's Signature <i>Antje Wiegand</i>	Date 28.05.01
Residence Jena		Citizenship German
Post Office Address Theo-Neubauer-Str. 10, D-07743 Jena Germany		

DEX

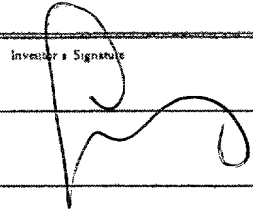
Full Name of Second Joint Inventor, if any	Inventor's Signature	Date
Residence		Citizenship
Post Office Address		

Full Name of Sole or First Inventor	Inventor's Signature	Date
Residence		Citizenship
Post Office Address		

Full Name of Second Joint Inventor, if any	Inventor's Signature	Date
Residence		Citizenship
Post Office Address		

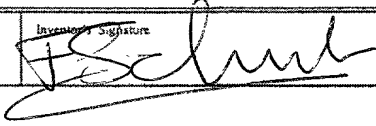
Full Name of Second Joint Inventor, if any	Inventor's Signature	Date
Residence		Citizenship
Post Office Address		

5-00

Full Name of Sole or First Inventor <u>Louis OOSTING</u>	Inventor's Signature 	Date <u>01-06-2001</u>
Residence <u>Groningen</u>	Citizenship <u>Dutch</u>	
Post Office Address <u>Helper Esweg 23, NL-9722 RP Groningen</u>		<u>Netherlands</u>

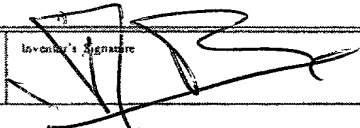
NLX

6-00

Full Name of Second Joint Inventor, if any <u>Frederik SCHUT</u>	Inventor's Signature 	Date <u>01.06.2001</u>
Residence <u>Groningen</u>	Citizenship <u>Dutch</u>	
Post Office Address <u>J.H. Diemer Straat 25, NL-9728 VM Groningen</u>		<u>Netherlands</u>

NLX

7-00

Full Name of Second Joint Inventor, if any <u>Paris, Som, Tjwan Tan</u>	Inventor's Signature 	Date <u>01-06-01</u>
Residence <u>Haren</u>	Citizenship <u>Dutch</u>	
Post Office Address <u>Emdaborg Nr. 14, NL-9751 SJ Haren,</u>		<u>Netherlands</u>

NLX

Full Name of Sole or First Inventor	Inventor's Signature	Date
Residence	Citizenship	
Post Office Address		

Full Name of Second Joint Inventor, if any	Inventor's Signature	Date
Residence	Citizenship	
Post Office Address		

Full Name of Second Joint Inventor, if any	Inventor's Signature	Date
Residence	Citizenship	
Post Office Address		